Designation: WIL2

CLS order number: Cryovial: 30201

Cryovial: 302011 DNA: 302011GD



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Origin and General Cha	racteristics	
Organism:	Homo sapiens	
Tissue:	Hematopoietic system	
Morphology:	B-Lymphoblast	
Growth Properties:	Suspension	
Description:	Derived from the spleen of a 5-year-old Caucasian male with hereditary spherocytosis. Cell line contains Epstein Barr-Virus. Further information for the cell line see Cellsaurus, Acession CVCL_6544 A variant thereof (Wi-L2-HF2v or WI-L2-729HF2) was used as a fusion partner for the production of human hybridomas (US patent US4594325A, June 10,1986, application status: expired (2019).	
References:	Levy J.A., Virolainen M., Defendi V. Human lymphoblastoid lines from lymph node and spleen. Cancer 22:517-524(1968) Levy J.A., Buell D.N., Creech C., Hirshaut Y., Silverberg H. Further characterization of the WI-L1 and WI-L2 lymphoblastoid lines. J. Natl. Cancer Inst. 46:647-654(1971) Stiles C.D., Desmond W. Jr., Sato G., Saier M.H. Jr. Failure of human cells transformed by simian virus 40 to form tumors in athymic nude mice. Proc. Natl. Acad. Sci. U.S.A. 72:4971-4975(1975) Uittenbogaart C.H., Cantor Y., Fahey J.L. Growth of human malignant lymphoid cell lines in serum-free medium. In Vitro 19:67-72(1983) Evans H.H., Ricanati M., Horng MF., Jiang Q., Mencl J., Olive P. DNA double-strand break rejoining deficiency in TK6 and other human B-lymphoblast cell lines. Radiat. Res. 134:307-315(1993)	
Culture Conditions and	Handling	
Culture Medium:	RPMI Advanced supplemented with glucose, Non-Essential Amino Acids, and sodium pyruvate. No FBS was added. (MG-74a, CLS order number 820704a).	
Subculturing:	Maintain culture between 3 to 9 x10 ⁵ cells/ml; A maximum density of 2 x10 ⁶ cells/ml is possible. Incubate at 5% CO ₂ , 37°C.	
Seeding density:	1x10 ⁵ /ml	
Fluid Renewal:	2 times weekly	
Freeze Medium:	CM-1 (CLS order number: 800125, 25ml, 800150, 50ml)	
Freezing recovery:	Fast	
Sterility:	Fluorescence (DAPI) test: negative; Mycoplasma specific PCR: negative; Bacteria specific PCR: negative	
Biosafety Level:	2	
Safety precautions:	If the cryovial is planned to be stored in liquid nitrogen and to be thawed in the future, special safety precautions should be followed: Protective gloves and clothing should be used and a facemask or safety goggles must be worn when transferring frozen samples into or removing from the liquid nitrogen tank. The removal of a cryovial from liquid nitrogen may result in the explosion of the frozen vial creating flying fragments. Caputo, J.L. Biosafety procedures in cell culture. J. Tissue Cult. Methods 11:223-227, 1988. ATCC Quality Control Methods for Cell Lines, 2nd edition, 1992.	
Special Features of the Cell Line		
Karyotype:	46, hypodiploid	
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DNA Profile (STR) CLS in accordance with STR-pattern published by Cellsaurus	Amelogenin: X,Y CSF1PO: 11,12 D13S317: 11 D16S539:11,12 D5S818: 12,13 D7S820:9,11 TH01: 8,9.3 TPOX: 8,11 D19S433: 14,16	vWA: 17,20 D3S1358: 16 D21S11: 29 D18S51:11,16 D8S1179: 10,13 FGA: 22,24 D2S1338: 17,25
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Certificate of Analysis: The Certificate of Analysis for each batch can be requested by e-mail at service@clsgmbh.de.

Recommendations for handling of cells growing in suspension following delivery			
Cryopreserved cells	The cells come deep-frozen shipped on dry ice. Please make sure that the vial is still frozen.		
	If immediate culturing is not intended, the cryovial(s) must be stored below -150°C after arrival.		
	If immediate culturing is intended, please follow these instructions:		
	Quickly thaw by rapid agitation in a 37°C water bath within 40-60 seconds. The water bath should have clean water containing an antimicrobial agent. As soon as the sample has thawed, remove the cryovial from the water bath. Note: A small ice clump should still remain and the vial should still be cold.		
	From now on, all operations should be carried out under aseptic conditions.		
	Transfer the cryovial to a sterile flow cabinet and wipe with 70% alcohol. Carefully open the vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of culture medium (room temperature). Resuspend the cells carefully. Centrifuge at 300xg for 3 min and discard the supernatant. The centrifugation step may be omitted, but in this case the remains of the freeze medium have to be removed 24 hours later.		
	Resuspend the cells carefully in 10ml fresh cell culture medium and transfer them into one T25 cell culture flask. All further steps are described in the Subculture section.		
Proliferating Cultures	The cell culture flask, 1xT25, comes filled with cell culture medium. Incubate at 37°C for a minimum of 24 hrs.		
	Count the cells, spin down the cell suspension at 300x g for 3 minutes to collect the cells. Resuspend the cells in an appropriate amount of fresh cell culture medium and transfer to new cell culture flasks.		
	Incubate at 37°C for a minimum of 24 hrs.		

Warranty:	CLS warrants for a high cell viability and culture performance only if the product(s) is (are) stored and cultured according to the information described above. Using cell culture media and supplements other than the ones recommended in this product information may result in satisfactory proliferation and viabilities. CLS, however, does not warrant for cell recovery, proliferation and function if differing formulations are employed.
Disclaimer:	The customer shall not be entitled to employ this product for purposes other than research. Commercial utilization shall not be permitted; in particular, the cell line, its components or materials made therefrom shall not be sold or transferred to any third party. In addition, the term 'Commercial use' shall mean any activity by a party for consideration and may include, but is not limited to, use of the product or its components in manufacturing, for providing services, e.g. fee for service testing, in quality control or assurance processes within the manufacturing of products for sale, for therapeutic, diagnostic or prophylactic purposes, or for resale.