

## **DATASHEET**

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## Lipopolysaccharides (LPS) Antibody Pair

Catalogue No.:abx370829

**Detection Antibody Clonality:** 

Capture Antibody Conjugation:

Lipopolysaccharide (LPS) Antibody Pair for use in Sandwich ELISA assay development. The 10 × 96 tests size contains:

- 400 µg Apolipoprotein A4 (APOA4) mouse monoclonal capture antibody,
- 30 µg Lipopolysaccharide (LPS) biotin-conjugated rabbit polyclonal detection antibody,
- 600 ng Lipopolysaccharide (LPS) standard.

It is recommended to use this antibody pair with abx098958 Antibody Pair Support Kit (Sandwich Method).

Target:	Lipopolysaccharide (LPS)
Reactivity:	General
Tested Applications:	ELISA
Recommended dilutions:	Dilute the Capture Antibody 210-fold with Coating Buffer.  Dilute the biotin-conjugated Detection Antibody 1100-fold with Detection Antibody Diluent.  Optimal dilutions/concentrations should be determined by the end user.
Form:	Liquid (Capture Antibody and Detection Antibody)
Reconstitution:	Reconstitute the standard with Standard Diluent. The volume, and therefore standard concentration, should be determined by the end user.
Test Range:	0.47 ng/ml - 30 ng/ml
Storage:	Store at 2 to 8 °C for up to one month. Aliquot and store at -80 °C for up to one year. Avoid repeated freeze/thaw cycles. All solutions should be made fresh before the experiment.
Concentration:	400 $\mu$ g/0.6 ml (Capture Antibody), 30 $\mu$ g/0.09 ml (Detection Antibody), 600 ng (Standard)
Standard Form:	Lyophilized
ELISA Type:	Sandwich
Capture Antibody Host:	Mouse
Detection Antibody Host:	Rabbit
Capture Antibody Clonality:	Monoclonal

Polyclonal

Unconjugated



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**Detection Antibody Conjugation: Biotin** 

**Buffer:** The capture and detection antibody both contain 0.1% sodium azide.

Directions for use: Bring all components to room temperature (18-25°C) and briefly spin or centrifuge the vials before use. Working solutions should be prepared and used immediately.

## Recommended Procedure:

- 1. Dilute the Capture Antibody to working concentration using Coating Buffer. Immediately coat the 96-well plate with diluted Capture Antibody (100 µl per well). Seal the plate and incubate at 4 °C overnight or at 37 °C for 2 hours
- 2. Aspirate the wells and wash with Wash Buffer (350 µl per well) and allow to soak for 1-2 min. Remove the liquid by inverting and tapping the plate on to absorbent paper.
- 3. Block the plate with Blocking Buffer (200 µl per well) at 37 °C for 1.5 hours.
- 4. Repeat the aspiration/wash process in Step 2.
- 5. Add 100 µl of standards or sample into the appropriate wells. Cover with a plate sealer and incubate at 37 °C for 1 hour.
- 6. Repeat the aspiration/wash process in Step 2.
- 7. Add appropriately diluted biotin-conjugated Detection Antibody (100 µl per well). Cover the plate with a new plate sealer and incubate at 37 °C for 1 hour.
- 8. Repeat the aspiration/wash process in Step 2.
- 9. Add appropriately diluted Streptavidin HRP (100 µl per well). Cover the plate with a new plate sealer and incubate at 37 °C for 30 min.
- 10. Repeat the aspiration/wash process in Step 2.
- 11. Add Substrate Solution (90 µl per well). Cover the plate with a new plate sealer and incubate at 37 °C for 10-20 min. Keep the plate in the dark and avoid exposure to light.
- 12. Add Stop Solution (50 µl per well). Tap the side of the plate to ensure thorough mixing.
- 13. Measure the absorbance immediately using a microplate reader set at 450 nm.

This product is for research use only.

Note: