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DATASHEET

Inter Alpha-Globulin Inhibitor H2 (ITIH2) Antibody Pair

Catalogue No.:abx370449

Inter Alpha-Globulin Inhibitor H2 (ITIH2) Antibody Pair for use in Sandwich ELISA assay development. This antibody pair contains:

- Inter Alpha-Globulin Inhibitor H2 (ITIH2) capture antibody,
- Inter Alpha-Globulin Inhibitor H2 (ITIH2) biotin-conjugated detection antibody,
- Inter Alpha-Globulin Inhibitor H2 (ITIH2) standard.

It is recommended to use this antibody pair with abx098958 Antibody Pair Support Kit (Sandwich Method).

Target:	Inter Alpha-Globulin Inhibitor H2 (ITIH2)
Reactivity:	Human
Tested Applications:	ELISA
Recommended dilutions:	Dilute the Capture Antibody with Coating Buffer. Dilute the biotin-conjugated Detection Antibody with Detection Antibody Diluent. Optimal dilutions/concentrations should be determined by the end user.
Form:	Liquid (Capture Antibody and Detection Antibody)
Reconstitution:	Reconstitute the standard with Standard Diluent. The volume, and therefore standard concentration, should be determined by the end user.
Storage:	Store at 2 to 8 °C for up to one month. Aliquot and store at -80 °C for up to one year. Avoid repeated freeze/thaw cycles. All solutions should be made fresh before the experiment.
Standard Form:	Lyophilized
ELISA Type:	Sandwich
Capture Antibody Conjugation:	Unconjugated
Detection Antibody Conjugation:	Biotin
Buffer:	The capture and detection antibody both contain 0.1% sodium azide.

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Directions for use:

Bring all components to room temperature (18-25°C) and briefly spin or centrifuge the vials before use. Working solutions should be prepared and used immediately.

Recommended Procedure:

1. Dilute the Capture Antibody to working concentration using Coating Buffer. Immediately coat the 96-well plate with diluted Capture Antibody (100 µl per well). Seal the plate and incubate at 4 °C overnight or at 37 °C for 2 hours
2. Aspirate the wells and wash with Wash Buffer (350 µl per well) and allow to soak for 1-2 min. Remove the liquid by inverting and tapping the plate on to absorbent paper.
3. Block the plate with Blocking Buffer (200 µl per well) at 37 °C for 1.5 hours.
4. Repeat the aspiration/wash process in Step 2.
5. Add 100 µl of standards or sample into the appropriate wells. Cover with a plate sealer and incubate at 37 °C for 1 hour.
6. Repeat the aspiration/wash process in Step 2.
7. Add appropriately diluted biotin-conjugated Detection Antibody (100 µl per well). Cover the plate with a new plate sealer and incubate at 37 °C for 1 hour.
8. Repeat the aspiration/wash process in Step 2.
9. Add appropriately diluted Streptavidin HRP (100 µl per well). Cover the plate with a new plate sealer and incubate at 37 °C for 30 min.
10. Repeat the aspiration/wash process in Step 2.
11. Add Substrate Solution (90 µl per well). Cover the plate with a new plate sealer and incubate at 37 °C for 10-20 min. Keep the plate in the dark and avoid exposure to light.
12. Add Stop Solution (50 µl per well). Tap the side of the plate to ensure thorough mixing.
13. Measure the absorbance immediately using a microplate reader set at 450 nm.

Note:

This product is for research use only.