

Chicken Anti-SRBC IgG ELISA

Life Diagnostics, Inc., Catalog Number: SRBCG-5

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INTRODUCTION

Measurement of anti-SRBC (sheep red blood cell) immunoglobulin levels is used to assess immune function in chickens. To date, a somewhat qualitative hemagglutination assay has often been used to measure antibody titers. Disadvantageously, the hemagglutination assay does not readily differentiate between IgG (IgY) and IgM responses. To address these issues, we at Life Diagnostics, Inc. have developed an enzyme linked immunosorbent assay (ELISA) that allows rapid and quantitative measurement of chicken anti-SRBC IgG levels in serum or plasma. We also manufacture a companion ELISA (catalog number SRBCM-5) that allows quantitative measurement of chicken anti-SRBC IgM levels.

Studies at Life Diagnostics, Inc. have demonstrated that IV immunization of chickens with 0.5 ml of a 10% solution of SRBC caused an elevation of anti-SRBC IgG levels from undetectable levels on day zero to ~11,000 units/ml on day seven.

PRINCIPLE OF THE ASSAY

The chicken anti-SRBC IgG ELISA uses detergent solubilized SRBC ghosts for solid phase (microtiter wells) immobilization and horseradish peroxidase (HRP) conjugated anti-chicken IgG antibodies for detection. Test serum or plasma samples are diluted and incubated in the microtiter wells for 45 minutes. The microtiter wells are subsequently washed, and HRP conjugate is added and incubated for 45 minutes. Anti-SRBC IgG molecules are thus sandwiched between immobilized SRBC antigens and the detection antibody conjugate. The wells are then washed to remove unbound HRP-labeled antibodies, and TMB Reagent is added and incubated for 20 minutes at room temperature. This results in the development of a blue color. Color development is stopped by the addition of Stop Solution, changing the color to yellow, and optical density is measured spectrophotometrically at 450 nm. The concentration of anti-SRBC IgG is proportional to the optical density of the test sample and is derived from a standard curve.

MATERIALS AND COMPONENTS

Materials provided with the kit:

- SRBC Coated 96-well Plate (provided as 12 strips of 8 wells)
- Enzyme Conjugate Reagent, 11 ml
- Reference Standard Stock^A (lyophilized)
- 20x Wash Solution, 50 ml
- Diluent, 50 ml
- TMB Reagent (One-Step), 11 ml
- Stop Solution (1N HCl), 11 ml

Materials required but not provided:

- Precision pipettes and tips
- Distilled or deionized water
- Polypropylene or glass tubes
- Vortex mixer
- Absorbent paper or paper towels
- Micro-plate incubator/shaker with mixing speed of ~150 rpm

- Plate washer
- Plate reader with an optical density range of 0-4 at 450nm
- Graph paper (PC graphing software is optional)

STORAGE

The kit should be stored at 2-8°C and the microtiter plate should be kept in a sealed bag with desiccant to minimize exposure to damp air. Test kits will remain stable for six months from the date of purchase provided that the components are stored as described above.

GENERAL INSTRUCTIONS

1. Please read and understand the instructions thoroughly before using the kit.
2. All reagents should be allowed to reach room temperature (18-25°C) before use.
3. The assay was designed for use with serum or plasma obtained from chickens 5-7 days after immunization with SRBC.
4. The optimal sample dilution should be determined empirically. However, studies performed at Life Diagnostics, Inc. suggest an initial sample dilution of 500 fold.
5. Optimal results are achieved if, at each step, reagents are pipetted into the wells of the microtiter plate within 5 minutes.

WASH SOLUTION PREPARATION

The wash solution is provided as a 20x stock. Prior to use, dilute the contents of the bottle (50 ml) with 950 ml of distilled or deionized water.

STANDARD PREPARATION

1. Reconstitute the vial of the lyophilized chicken anti-SRBC IgG standard stock with 200 µl of diluents, and then prepare the 100 u/ml standard in a polypropylene or glass tube as described on the vial label.
2. Label 4 polypropylene or glass tubes as 50, 25, 12.5, and 6.25 u/ml, and dispense 250 µl of diluent into each tube.
3. Prepare a 50 u/ml standard by diluting and mixing 250 µl of the 100 u/ml standard with 250 µl of diluent in the tube labeled 50 u/ml.
4. Similarly prepare the 25, 12.5 and 6.25 u/ml standards by serial dilution.

After reconstitution the standard stock solution is stable for at least one week if stored at 4°C. It should be frozen at or below -20°C if use beyond this time is intended. Avoid multiple freeze-thaws.

SAMPLE PREPARATION

General Note: Studies at Life Diagnostics, Inc. revealed anti-SRBC IgG levels of approximately 10,000 u/ml 7 days after immunization with SRBC. In order to obtain values within the range of the standard curve, we suggest that samples initially be diluted 500 fold using the following procedure:

1. For each test sample dispense 90 µl and 200 µl of diluent into separate tubes.

^A Chicken anti-SRBC IgG levels are measured in nominal units and are calibrated with reference to chicken anti-SRBC serum prepared at Life Diagnostics, Inc.

2. Pipette and mix 10 μ l of the serum/plasma sample into the tube containing 90 μ l of diluent. This provides a 10 fold diluted sample.
3. Mix 5 μ l of the 10-fold diluted sample with 245 μ l of diluent in the second tube to give a 500-fold dilution of the sample.
4. Repeat this procedure for each sample to be tested.

ASSAY PROCEDURE

1. Secure the desired number of coated wells in the holder.
2. Dispense 100 μ l of standards and diluted samples into the wells (we recommend that samples and standards be tested in duplicate).
3. Incubate on an orbital micro-plate shaker at 100-150 rpm at room temperature (18-25°C) for 45 minutes.
4. Aspirate the contents of the microtiter wells and wash the wells 5 times with 1x wash solution using a plate washer (400 μ l/well). The entire wash procedure should be performed as quickly as possible.
5. Strike the wells sharply onto absorbent paper or paper towels to remove all residual wash buffer.
6. Add 100 μ l of enzyme conjugate reagent into each well.
7. Incubate on an orbital micro-plate shaker at 100-150 rpm at room temperature (18-25°C) for 45 minutes.
8. Wash as detailed in 4 to 5 above.
9. Dispense 100 μ l of TMB Reagent into each well.
10. Gently mix on an orbital micro-plate shaker at 100-150 rpm at room temperature (18-25°C) for 20 minutes.
11. Stop the reaction by adding 100 μ l of Stop Solution to each well.
12. Gently mix. *It is important to make sure that all the blue color changes to yellow.*
13. Read the optical density at 450 nm with a microtiter plate reader within 5 minutes.

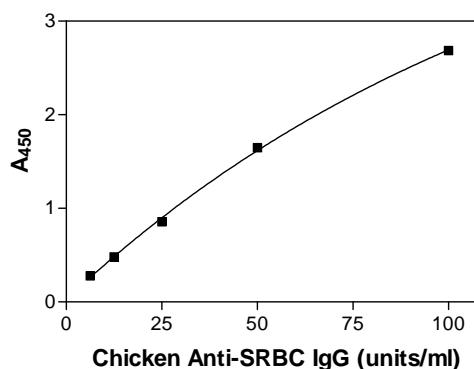
CALCULATION OF RESULTS

1. Calculate the average absorbance values (A_{450}) for each set of reference standards and samples.
2. Construct a standard curve by plotting the mean absorbance obtained from each reference standard against its concentration in u/ml on linear graph paper, with absorbance values on the vertical or Y-axis and concentrations on the horizontal or X-axis.
3. Using the mean absorbance value for each sample, determine the corresponding concentration of anti-SRBC IgG in u/ml from the standard curve.
4. Multiply the derived concentration by the dilution factor to determine the actual concentration of anti-SRBC IgG in the serum/plasma sample.
5. PC graphing software may be used for the above steps.
6. If the OD_{450} values of samples fall outside the standard curve when tested at a dilution of 500, samples should be diluted appropriately and re-tested.

TYPICAL STANDARD CURVE

A typical standard curve with optical density readings at 450nm on the Y-axis against anti-SRBC IgG concentrations on the X-axis is shown below. This curve is for the purpose of illustration only and should not be used to calculate unknowns. Each user should obtain his or her data and standard curve in each experiment.

Anti-SRBC IgG (u/ml)	A_{450}
100	2.686
50	1.649
25	0.855
12.5	0.481
6.25	0.279



LIMITATIONS OF THE PROCEDURE

1. Reliable and reproducible results will be obtained when the assay procedure is carried out with a complete understanding of and in accordance with the instructions detailed above.
2. The wash procedure is critical. Insufficient washing will result in poor precision and falsely elevated absorbance readings.

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For technical assistance please email us at
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