

1M DMSO And DAP213 (Media for Mouse Embryo Cytopreservation)

Cat. No. CSR-R-T073

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*Keep them at 4°C until use. Use all the media once opened and avoid using the remaining residue as it is not so stable for repetitive use.

Collection of embryos

Collect the embryos of the required stage by *in vitro* fertilization or oviduct perfusion after mating [Please refer to the datasheet of HTF (#CSR-R-B070, #CSR-R-B071) for *in vitro* fertilization, KSOM (#CSR-R-B074, #CSR-R-B075) or mWM (#CSR-R-B080, #CSR-R-B081) for oviduct perfusion after mating.]

Preparation In Prior to Use

1. Reconstitute P10 to room temperature.
2. Prepare cryotubes depending on the number of embryos you freeze.
3. Prepare to keep samples to 0°C with crushed ice, chiller, lab top cooler or other cooling devices and also cool DAP213 to 0°C .
4. Prepare cryoboxes or cryocanes according to the number of tubes you freeze. (They should be kept in liquid nitrogen)

Cryopreservation

1. Put DMSO drops into a dish. The number of drops should correspond to number of tubes you freeze + 1.
2. Place embryos to one of the drops and wait until the embryo drops down.
3. Divide the embryos equally and move them to each DMSO drops remaining with glass capillary.
4. Adjust a pipette to 5µL and transfer the embryos with DMSO into a cryotube.
5. Keep the cryotube in 0°C .
6. Add 45 µL of DMSO into the cryotube along the inner surface and equilibrate for 5 minute.
7. Put the tubes to pre-cooled cryobox or cryocanes, and preserve them in liquid nitrogen.
8. Keep them into liquid nitrogen to maintain frozen condition at all times in preservation. In case they are exposed to atmosphere and melt, it will lead to serious damage (survival rate will be remarkably decreased)



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