

# **RayBio® Biotin Label-based Human Antibody Array I**

**For the Simultaneous Detection of the Expression Levels of  
507 Human Proteins in Cell Culture Supernates.**

## **User Manual (Revised Apr 1, 2009)**

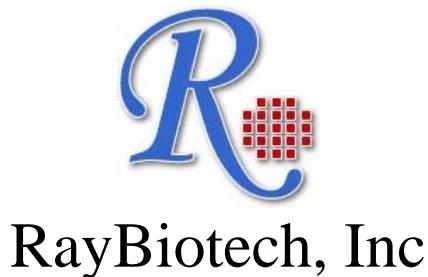
**( Cat#: AAH-BLM-1-2;  
AAH-BLM-1-4 )**



**As the Protein Array Pioneer Company,  
Excellence and Innovation Is Our Goal**

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RayBiotech, Inc

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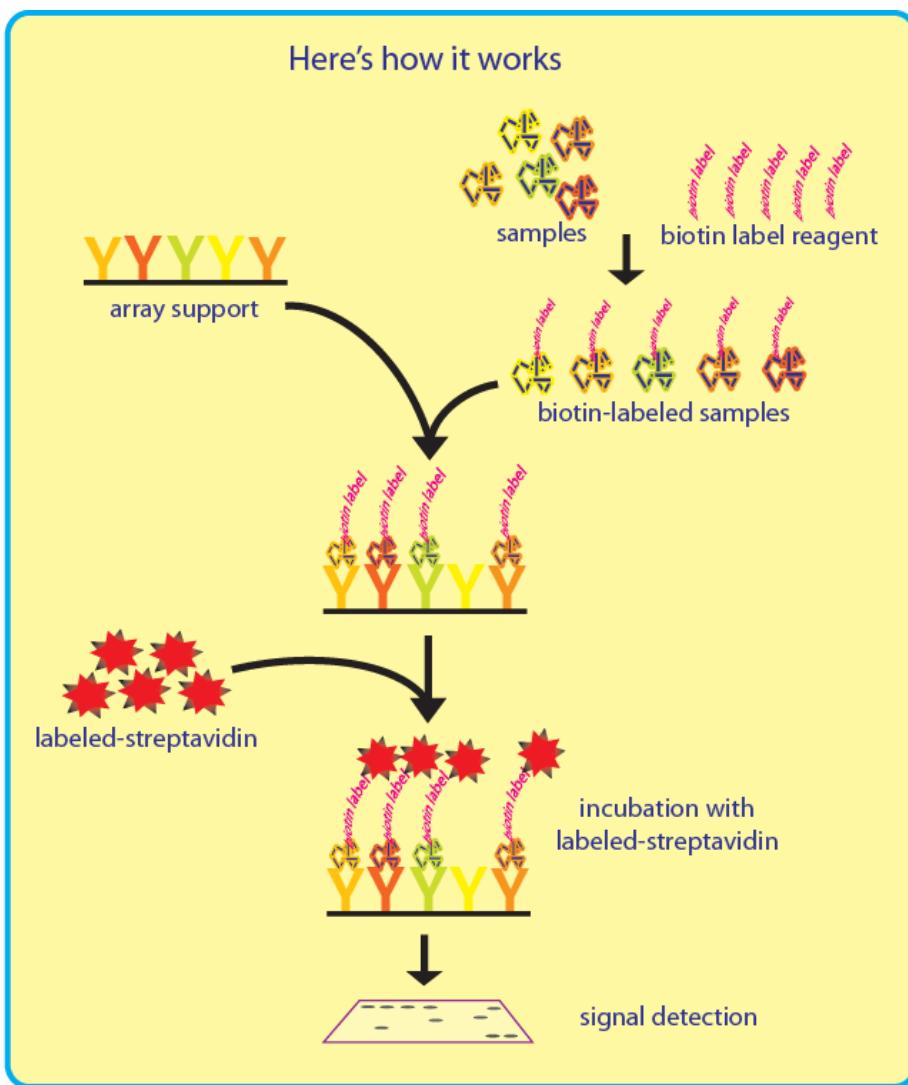
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## I. Introduction

Recent technological advances by Raybiotech have enabled the largest commercially available antibody array to date. With the L Series 507, researchers can now obtain a broad, panoramic view of cytokine expression. The expression levels of 507 human target proteins can be simultaneously detected, including cytokines, chemokines, adipokine, growth factors, angiogenic factors, proteases, soluble receptors, soluble adhesion molecules and other proteins in cell culture supernate and serum. Furthermore, an internal control is used to monitor the whole process including biotin-labeling, so this massive array will accurately reflect the available cytokines in your sample.

The first step in using the RayBio® Biotin label-based human antibody array 1 is to biotinylate the primary amine of the proteins in cell culture supernates. The biotin-labeled sample is then added onto array membrane and incubated at room temperature. After incubation with HRP-streptavidin, the signals can be visualized by chemiluminescence.



## II. Materials Provided

Upon receipt, the Box 1 should be stored at -20 °C and Box 2 should be stored at 4 °C. Please use within 6 months from the date of shipment. After initial use, the Blocking Buffer, Stop solution, HRP-Conjugated Streptavidin, Detection Buffer C and D should be stored

at 4 °C to avoid repeated freeze-thaw cycles. The Array Membrane and Internal Control should be kept at -20 °C.

**Box-1 (store at -20 °C):**

- Labeling Reagent (Item B, 1 tube for 2 array membranes , and 2 for 4 array membranes )
- Internal control (Item C, 1 tube for 2 array membranes , and 2 for 4 array membranes )
- Stop Solution (Item D, 50 µl)
- RayBio® Biotin label-based human antibody array I (Item E, 2 for Cat#: AAH-BLM-1-2, and 4 for Cat#: AAH-BLM-1-4)
- Blocking Buffer (Item F, 30 ml for each bottle, 2 bottles for 2 array membranes , and 4 for 4 array membranes)
- 500X HRP-Conjugated Streptavidin Concentrate (Item I, 100 µl)
- Detection Buffer C (Item K, 5 ml for 2 membranes, and 10 ml for 4 membranes)
- Detection Buffer D (Item L, 5 ml for 2 membranes, and 10 ml for 4 membranes)
- Plastic sheet

**Box 2 (store at 4 °C):**

- Dialysis tube and Floating Rack (Item A, 2 tubes for 2 array membranes, and 4 for 4 array membranes)
- 20X Wash Buffer I (Item G, 30ml)
- 20X Wash Buffer II (Item H, 30ml)
- Spin Column (Item J, 2 columns for 2 array membranes, and 4 for 4 array membranes)
- Plate (2 plates for 2 array membranes, and 4 for 4 array membranes)

### **III. Additional Materials Required**

- 1X PBS, pH=8.0
- Shaker
- 2~5 ml tube
- 50 ml conical collection tube
- Distilled water
- Kodak X-Omat™ AR film (REF 165 1454) and film processor or Chemiluminescence imaging system

### **IV. Overview and General Considerations**

#### **A. Handling Array Membranes**

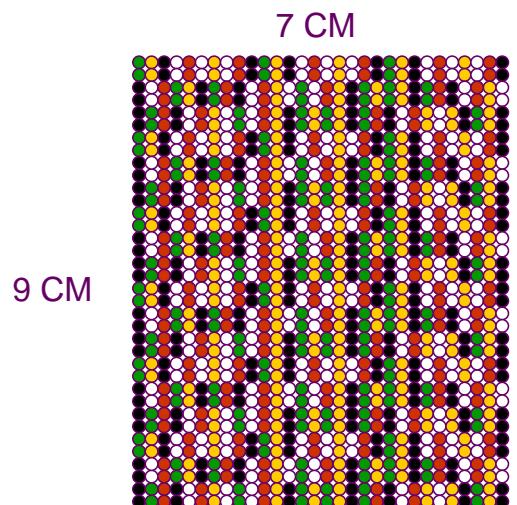
- Always use forceps to handle membranes, and grip the membranes by the edges only.
- Never allow array membranes to dry during experiments.
- Avoid touch Array membrane by hand, tips or any sharp tools.

#### **B. Incubation**

- Completely cover membranes with sample or buffer during incubation, and cover eight-well tray with lid to avoid drying.
- Avoid foaming during incubation steps.
- Perform all incubation and wash steps under gentle rotation.
- Several incubation steps such as step 3 in page 10 (sample incubation) or step 7 in page 11 (HRP-streptavidin incubation) may be done at 4 °C for overnight.

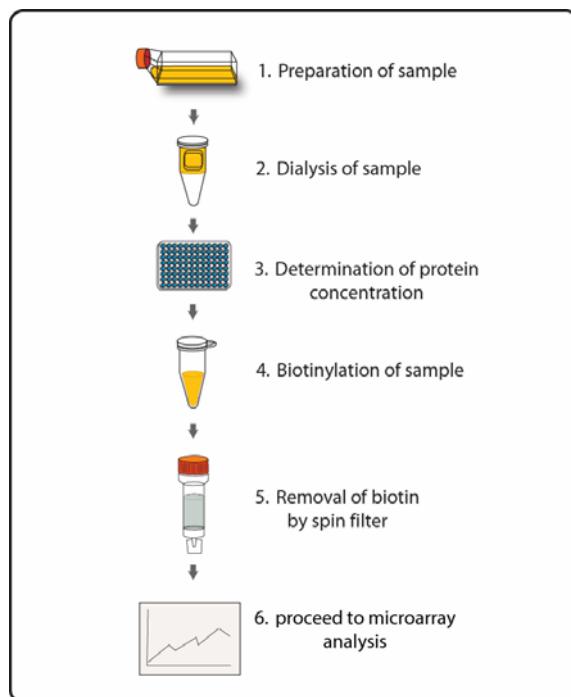
## V. Protocol

### Layout of Array Membrane



**30 columns x 36 rows**

### Assay Diagram



## **A. Preparation of Samples**

The cell culture supernates can be prepared in the following conventional manner:

To prepare cell culture supernates (cell conditioned media), cells are plated in 100 mm tissue culture dishes at a density of  $1 \times 10^6$  cells\* per dish. The cells are then cultured with complete culture medium for 24~48 hours\*\*. The complete culture medium is replaced with lower serum medium such as 0.2% FCS serum, and then the cells are cultured for 48 hour\*\* again once more. The supernates are collected, centrifuged at 1,000 g, aliquoted and stored at  $-80^{\circ}\text{C}$  until use. Meanwhile, the cells are also collected and the total protein concentration is determined. For each sample it is recommended that the concentration of the supernates and cell lysate (help nomalize different cell culture supernates) be determined using a total protein assay (BCA Protein Assay Kit, Pierce, Prod# 23227).

*Note: \* The density of cells per dish used is dependent on the cell type  
More or less cells may be used.*

*\*\* Culture times may vary depending on your cell lines and research.*

## **B. Dialysis of Sample**

The cell culture supernates should be dialyzed with a Dialysis tube (Item A) before the biotin-labeling procedure. We recommend loading 2.5~3.0 ml cell culture supernates into a dialyzer and dialyzing with at least 2,000 ml 1X PBS buffer (pH = 8) at  $4^{\circ}\text{C}$ . Change the 1X PBS buffer and dialyze again. Allow at

least 3 h for each dialysis step, stir gently. The sample total volume may be changed after dialysis.

*Note: Preparation of 1X PBS, pH=8.0, 1.0 g KCl, 40 g NaCl, 1.0 g KH<sub>2</sub>PO<sub>4</sub>, 5.75 g Na<sub>2</sub>HPO<sub>4</sub> dissolve in 4,500 ml deionized or distilled water. Adjust pH=8.0 with 1M NaOH and adjust final volume to 5,000 ml with deionized or distilled water.*

### **C. Biotin-labeling Sample**

Avoid contamination with any solution containing amines (i.e., Tris, glycine) as well as Azide during the biotinylation process.

1. Briefly spin down Internal Control tube (Item C) before use. Add 100 µl 1X PBS, pH=8.0 into the Internal Control tube, pipette up and down to dissolve the powder. Transfer 2 ml dialyzed sample into a new tube. Add 40 µl prepared Internal Control into the tube. Mix well.
2. Immediately before use, briefly spin down the Labeling Reagent tube (Item B). Add 100 µl 1X PBS into the tube, pipette up and down or vortex to dissolve the powder to prepare 1X Labeling Reagent solution.
3. Add an appropriate amount\* of prepared Labeling Reagent into above tube with sample in step 2, mix well immediately. Incubate the reaction solution at room temperature for 30 min with gentle shaking. Gently tap the tube to mix the reaction solution every 5 min.

- \* 7.2  $\mu$ l of 1X Labeling Reagent for labeling 1 mg total protein in supernates .

*Note: You need to re-calculate the total protein concentration if cell culture supernatet volume is changed after dialysis and you measure the total protein concentration before dialysis step.*

4. Add 5  $\mu$ l Stop Solution into the above reaction solution and then use the spin column to remove free biotin.
  - a). Twist off the spin column's bottom closure and loosen the cap. Place the column into a 50 ml collection tube.
  - b). Centrifuge column at 1,000 g for 3 minutes to remove storage solution.

*Note: The resin will appear compacted after centrifugation.*

- c). Add 5 ml 1X PBS into column, centrifuge at 1,000 g for 3 minutes to 1X PBS. Repeat additional 2 times to wash the column.
- d). Place the column in a new collection tube, slowly load the sample to the center of the compact resin bed.
- e). Centrifuge the column at 1,000 g for 3 minutes to collect sample. Stored at –80  $^{\circ}$ C until testing. Discard column after use.

## **D. Blocking and Incubation**

1. Place each membrane into the provided tray (“-” mark is on the antibody printed side).

***Note:*** *The printed side should be facing upward.*

2. Add 8 ml Blocking Buffer and incubate at room temperature with gentle shaking for 1 hour to block membranes.
3. Decant Blocking Buffer from each container. Add 8 ml of sample into each array membrane, and cover with the lid. Incubate at room temperature with gentle shaking for 2 hours. Dilute sample using Blocking Buffer.

***Note:*** *1). We recommended using 8 ml of 5-fold diluted cell culture supernates which have been biotin-labeled. Dilute sample using Blocking Buffer.*

***Note:*** *2). The amount of sample used depends on the abundance of protein. More of the sample can be used if signals are too weak. If signals are too strong, the sample can be diluted further.*

***Note:*** *3). Incubation may be done at room temperature for 2 hours. Over night at 4°C*

4. Decant the samples from each container, and wash 3 times with 20 ml of 1X Wash Buffer I at room temperature with shaking. 5

- min per wash. Dilute 20X Wash Buffer I with deionized or distilled water.
5. Decant the 1X Wash Buffer I from each container. Wash 3 times with 20 ml of 1X Wash Buffer II at room temperature.
  6. Decant the 1X Wash Buffer II. Add 8 ml of 500 fold diluted HRP-conjugated streptavidin (e.g. add 36 µl of HRP-conjugated streptavidin to 18 ml of Blocking Buffer) to each membrane.

*Note: Mix tube containing 500X HRP-Conjugated Streptavidin well before use since precipitation may form during storage.*
  7. Incubate at room temperature with gentle shaking for 2 hours.

*Note: incubation may be done at 4°C for overnight.*
  8. Wash as directed in steps 4 and 5.

## **E. Detection**

- \* Do not let the membrane dry out during detection. The detection process must be completed within 40 minutes without stopping.**
1. Add 4.2 ml of Detection Buffer C and 4.2 ml of Detection Buffer D into a tube (for detecting 2 membranes); Mix both solutions; Drain off excess wash buffer. Place membrane protein side up (“-” mark is on the protein side top left corner) on a clean plastic plate or its cover (provided in the kit). Pipette 4ml of the mixed

Detection Buffer on to each membrane and incubate at room temperature for 2 minutes with shaking. Ensure that the detection mixture is evenly covering the membrane without any air bubbles.

2. Gently place the membrane with forceps, protein side up, on a piece of plastic sheet (“-” mark is on the protein side top left corner). Cover the array with another piece of plastic sheet. Gently smooth out any air bubbles. Avoid using pressure on the membrane. Work as quickly as possible.
3. The signal can be detected directly from the membrane using a chemiluminescence imaging system or by exposing the array to x-ray film (we recommend using Kodak X-Omat™ AR film) with subsequent development. Expose the membranes for 40 Seconds. Then re-expose the film according to the intensity of signals. If the signals are too strong (background too high), reduce exposure time (eg, 5–30 seconds). If the signals are too weak, increase exposure time (eg, 5–20 min or overnight). Or re-incubate membranes overnight with 1X HRP-conjugated streptavidin, and repeat detection on the second day.
4. Save membranes at –20 °C to –80 °C for future reference.

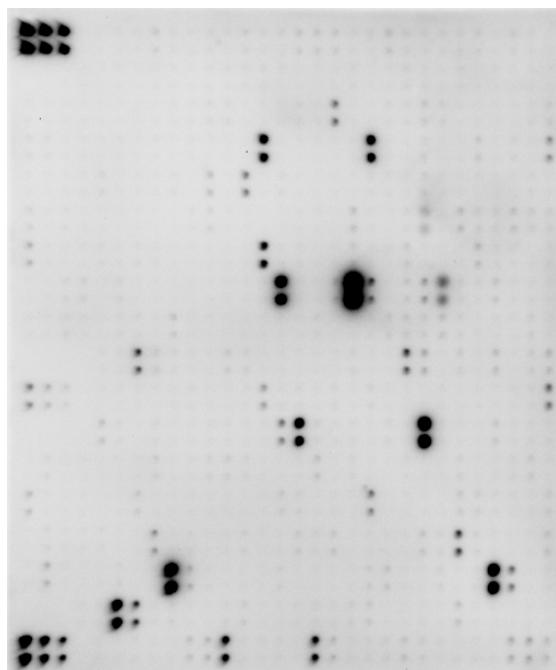
## **VI. Interpretation of Results:**

The following figure shows the RayBio® **Biotin-label-based Array I** probed with cell culture supernates. The image was captured using a chemiluminescence imaging system. One important parameter is the background signal. To obtain the best

results, we suggest that several exposures be attempted. By comparing the signal intensities, relative expression levels of target proteins can be made. The intensities of signals can be quantified by densitometry. A biotinylated protein and internal control will produce positive control signals, which can be used to identify the orientation and help normalize the results from different arrays being compared.

Antibody affinity to its target varies significantly between antibodies. The intensity detected on the array with each antibody depends on this affinity; therefore, signal intensity comparison can be performed only within the same antibody/antigen system and not between different antibodies.

The **RayBio® Analysis Tool** is a program specifically designed for analysis of RayBio® Biotin Label-based Antibody Arrays. This tool will not only assist in compiling and organizing your data, but also reduces your calculations to a “copy and paste.” Call RayBiotech, Inc. at 770-729-2992 for ordering information.



RayBio® Biotin label-based Array I Protocol

**RayBio® Biotin Label-based Human Antibody Array I Map –**  
**Larger versions of this can obtained by contacting technical support at 770-729-2992.**

P-1a	P-1b	P-1c	N	5	6	7	8	9	10	11	12	13	14	15	16	17	18	19	20	21	22	23	24	25	26	27	28	29	30	
P-1a	P-1b	P-1c	N	5	6	7	8	9	10	11	12	13	14	15	16	17	18	19	20	21	22	23	24	25	26	27	28	29	30	
N	N	N	34	35	36	37	38	39	40	41	42	43	44	45	46	47	48	49	50	51	52	53	54	55	56	57	58	59	60	
61	62	63	64	65	66	67	68	69	70	71	72	73	74	75	76	77	78	79	80	81	82	83	84	85	86	87	88	89	90	
61	62	63	64	65	66	67	68	69	70	71	72	73	74	75	76	77	78	79	80	81	82	83	84	85	86	87	88	89	90	
91	92	93	94	95	96	97	98	99	100	101	102	103	104	105	106	107	108	109	110	111	112	113	114	115	116	117	118	119	120	
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151	152	153	154	155	156	157	158	159	160	161	162	163	164	165	166	167	168	169	170	171	172	173	174	175	176	177	178	179	180	
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B	B	B	B	275	276	277	278	279	280	281	282	283	284	285	286	287	288	289	290	291	292	293	294	295	296	297	298	299	300	
B	B	B	B	275	276	277	278	279	280	281	282	283	284	285	286	287	288	289	290	291	292	293	294	295	296	297	298	299	300	
IC-1a	IC-1b	IC-1c	B	305	306	307	308	309	310	311	312	313	314	315	316	317	318	319	320	321	322	323	324	325	326	327	328	329	330	
IC-1a	IC-1b	IC-1c	B	305	306	307	308	309	310	311	312	313	314	315	316	317	318	319	320	321	322	323	324	325	326	327	328	329	330	
B	B	B	B	335	336	337	338	339	340	341	342	343	344	345	346	347	348	349	350	351	352	353	354	355	356	357	358	359	360	
B	B	B	B	335	336	337	338	339	340	341	342	343	344	345	346	347	348	349	350	351	352	353	354	355	356	357	358	359	360	
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B	B	B	B	484	485	486	487	488	489	490	491	492	493	494	495	496	497	498	499	500	501	502	503	504	505	506	507	N	N	N
P-2a	P-2b	P-2c	B	515	516	517	518	519	520	521	522	523	524	525	526	527	528	529	530	531	532	533	534	535	536	N	IC-2a	IC-2b	IC-2c	
P-2a	P-2b	P-2c	B	515	516	517	518	519	520	521	522	523	524	525	526	527	528	529	530	531	532	533	534	535	536	N	IC-2a	IC-2b	IC-2c	

# RayBio® Biotin Label-based Human Antibody List

1	Positive 1a	61	CCR4	121	EN-RAGE	181	GDF-15	241	IL-1 F10 / IL-1HY2
2	Positive 1b	62	CCR5	122	Eotaxin / CCL11	182	GDNF	242	IL-1 R3 / IL-1 R AcP
3	Positive 1c	63	CCR6	123	Eotaxin-2 / MPIF-2	183	GFR alpha-1	243	IL-1 R4 /ST2
4	neg	64	CCR7	124	Eotaxin-3 / CCL26	184	GFR alpha-2	244	IL-1 R6 / IL-1 Rrp2
5	6Ckine	65	CCR8	125	Epiregulin	185	GFR alpha-3	245	IL-1 R8
6	Activin A	66	CCR9	126	ErbB2	186	GFR alpha-4	246	IL-1 R9
7	Activin B	67	CD14	127	ErbB3	187	GITR / TNFRF18	247	IL-1 ra
8	Activin C	68	CD27 / TNFRSF7	128	ErbB4	188	GITR Ligand / TNFSF18	248	IL-1 sRI
9	Activin RIA / ALK-2	69	CD30 / TNFRSF8	129	Erythropoietin	189	Glucagon	249	IL-1 sRII
10	Activin RIB / ALK-4	70	CD30 Ligand / TNFSF8	130	E-Selectin	190	Glut1	250	IL-2
11	Activin RII A/B	71	CD40 / TNFRSF5	131	ETL	191	Glut2	251	IL-2 R alpha
12	Activin RIIA	72	CD40 Ligand / TNFSF5 / CD154	132	FADD	192	Glut3	252	IL-2 R beta /CD122
13	Adiponectin / Acrp30	73	CD 163	133	FAM3B	193	Glut5	253	IL-2 R gamma
14	AgRP	74	Cerberus 1	134	Fas / TNFRSF6	194	Glypican 3	254	IL-3
15	ALCAM	75	Chem R23	135	Fas Ligand	195	Glypican 5	255	IL-3 R alpha
16	Angiogenin	76	Chordin-Like 1	136	FGF Basic	196	GM-CSF	256	IL-4
17	Angiopoietin-1	77	Chordin-Like 2	137	FGF-BP	197	GM-CSF R alpha	257	IL-4 R
18	Angiopoietin-2	78	Csk	138	FGF R3	198	Granzyme A	258	IL-5
19	Angiopoietin-4	79	CLC	139	FGF R4	199	GREMLIN	259	IL-5 R alpha
20	Angiopoietin-like 1	80	CNTF	140	FGF R5	200	GRO	260	IL-6
21	Angiopoietin-like 2	81	CNTF R alpha	141	FGF-4	201	GRO-a	261	IL-6 R
22	Angiopoietin-like Factor	82	Coagulation Factor III / Tissue	142	FGF-5	202	Growth Hormone (GH)	262	IL-7
23	Angiostatin	83	CRIM 1	143	FGF-6	203	Growth Hormone R (GHR)	263	IL-7 R alpha
24	APJ	84	Cripto-1	144	FGF-7 / KGF	204	HB-EGF	264	IL-8
25	AR (Amphiregulin)	85	CRTH-2	145	FGF-8	205	HCC-4 / CCL16	265	IL-9
26	APRIL	86	Cryptic	146	FGF-9	206	HCR / CRAM-A/B	266	IL-10
27	Artemin	87	CTACK / CCL27	147	FGF-10 / KGF-2	207	Hepassocin	267	IL-10 R alpha
28	AxL	88	CTGF / CCN2	148	FGF-11	208	Heregulin / NDF / GGF /	268	IL-10 R beta
29	B7-1 / CD80	89	CTLA-4 / CD152	149	FGF-12	209	HGF	269	IL-11
30	BAFF R / TNFRSF13C	90	CV-2 / Crossveinless-2	150	FGF-13 1B	210	HGFR	270	IL-12 p40
31	neg	91	CXCL14 / BRAK	151	FGF-16	211	HRG-alpha	271	Blank
32	neg	92	CXCL16	152	FGF-17	212	HRG-beta 1	272	Blank
33	neg	93	CXCR1 / IL-8 RA	153	FGF-18	213	HVEM / TNFRSF14	273	Blank
34	BCMA / TNFRSF17	94	CXCR2 / IL-8 RB	154	FGF-19	214	I-309	274	Blank
35	BD-1	95	CXCR3	155	FGF-20	215	ICAM-1	275	IL-12 p70
36	BDNF	96	CXCR4 (fusin)	156	FGF-21	216	ICAM-2	276	IL-12 R beta 1
37	beta-Catenin	97	CXCR5 / BLR-1	157	FGF-23	217	ICAM-3 (CD50)	277	IL-12 R beta 2
38	beta-Defensin 2	98	CXCR6	158	FLRG	218	ICAM-5	278	IL-13
39	beta-NGF	99	D6	159	Flt-3 Ligand	219	IFN-alpha / beta R1	279	IL-13 R alpha 1
40	BIK	100	DAN	160	Follistatin	220	IFN-alpha / beta R2	280	IL-13 R alpha 2
41	BLC / BCA-1 / CXCL13	101	DANCE	161	Follistatin-like 1	221	IFN-beta	281	IL-15
42	BMP-2	102	DcR3 / TNFRSF6B	162	Fractalkine	222	IFN-gamma	282	IL-15 R alpha
43	BMP-3	103	Decorin	163	Frizzled-1	223	IFN-gamma R1	283	IL-16
44	BMP-3b / GDF-10	104	Dkk-1	164	Frizzled-3	224	IGFBP-1	284	IL-17
45	BMP-4	105	Dkk-3	165	Frizzled-4	225	IGFBP-2	285	IL-17B
46	BMP-5	106	Dkk-4	166	Frizzled-5	226	IGFBP-3	286	IL-17B R
47	BMP-6	107	DR3 / TNFRSF25	167	Frizzled-6	227	IGFBP-4	287	IL-17C
48	BMP-7	108	DR6 / TNFRSF21	168	Frizzled-7	228	IGFBP-6	288	IL-17D
49	BMP-8	109	Dtk	169	Galectin-3	229	IGFBP-rp1 / IGFBP-7	289	IL-17E
50	BMP-15	110	EDA-A2	170	GASP-1 /	230	IGF-I	290	IL-17F
51	BMPR-IA / ALK-3	111	EDAR	171	GASP-2 / WFIKKN	231	IGF-I SR	291	IL-17R
52	BMPR-IB / ALK-6	112	EDG-1	172	GCP-2 / CXCL6	232	IGF-II	292	IL-17RC
53	BMPR-II	113	EGF	173	GCSF	233	IGF-II R	293	IL-17RD
54	BTC	114	EGF R / ErbB1	174	G-CSF R / CD 114	234	IL-1 alpha	294	IL-18 BPa
55	Cardiotrophin-1 / CT-1	115	EG-VEGF / PK1	175	GDF1	235	IL-1 beta	295	IL-18 R alpha /IL-1 R5
56	CCL14 / HCC-1 / HCC-3	116	EMAP-II	176	GDF3	236	IL-1 F5 / FIL1delta	296	IL-18 R beta /AcPL
57	CCL28 / VIC	117	ENA-78	177	GDF5	237	IL-1 F6 / FIL1 epsilon	297	IL-19
58	CCR1	118	Endocan	178	GDF8	238	IL-1 F7 / FIL1 zeta	298	IL-20
59	CCR2	119	Endoglin / CD105	179	GDF9	239	IL-1 F8 / FIL1 eta	299	IL-20 R alpha
60	CCR3	120	Endostatin	180	GDF11	240	IL-1 F9 / IL-1 H1	300	IL-20 R beta

# RayBio® Biotin Label-based Human Antibody List...continued

301	Internal Control 1a	361	MFRP	421	PECAM-1 / CD31	481	Blank
302	Internal Control 1b	362	MIF	422	Pentraxin3 / TSG-14	482	Blank
303	Internal Control 1c	363	MIG	423	Persephin	483	Blank
304	Blank	364	MIP-1a	424	PF4 / CXCL4	484	Tie-1
305	IL-21	365	MIP-1b	425	PIGF	485	Tie-2
306	IL-21 R	366	MIP-1d	426	PLUNC	486	TIMP-1
307	IL-22	367	MIP 2	427	Pref-1	487	TIMP-2
308	IL-22 BP	368	MIP-3 alpha	428	Progranulin	488	TIMP-3
309	IL-22 R	369	MIP-3 beta	429	Prolactin	489	TIMP-4
310	IL-23	370	MMP-1	430	P-selectin	490	TL1A / TNFSF15
311	IL-23 R	371	MMP-2	431	RAGE	491	TLR1
312	IL-24	372	MMP-3	432	RANK / TNFRSF11A	492	TLR2
313	IL-26	373	MMP-7	433	RANTES	493	TLR3
314	IL-27	374	MMP-8	434	RELM beta	494	TLR4
315	IL-28A	375	MMP-9	435	RELT / TNFRSF19L	495	TMEFF1 / Tomoregulin-1
316	IL-29	376	MMP-10	436	ROBO4	496	TMEFF2
317	IL-31	377	MMP-11 / Stromelysin-3	437	S100 A8/A9	497	TNF-alpha
318	IL-31 RA	378	MMP-12	438	S100A10	498	TNF-beta
319	Inhibin A	379	MMP-13	439	SAA	499	TNF RI / TNFRSF1A
320	Inhibin B	380	MMP-14	440	SCF	500	TNF RII / TNFRSF1B
321	Insulin	381	MMP-15	441	SCF R / CD117	501	TRADD
322	Insulin R	382	MMP-16 / MT3-MMP	442	SDF-1 / CXCL12	502	TRAIL / TNFSF10
323	Insulysin / IDE	383	MMP-19	443	sFRP-1	503	TRAIL R1 / DR4 / TNFRSF10A
324	IP-10	384	MMP-20	444	sFRP-3	504	TRAIL R2 / DR5 / TNFRSF10B
325	I-TAC / CXCL11	385	MMP-24 / MT5-MMP	445	sFRP-4	505	TRAIL R3 / TNFRSF10C
326	Kininostatin / kininogen	386	MMP-25 / MT6-MMP	446	sgp130	506	TRAIL R4 / TNFRSF10D
327	Kremen-1	387	Musk	447	SIGIRR	507	TRANCE
328	Kremen-2	388	MSP alpha Chain	448	Siglec-5/CD170	508	Neg
329	Lck	389	MSP beta-chain	449	Siglec-9	509	Neg
330	Latent TGF-beta bp1	390	NAP-2	450	SLPI	510	Neg
331	Blank	391	NCAM-1 / CD56	451	Smad 1	511	Positive 2a
332	Blank	392	Neuritin	452	Smad 4	512	Positive 2b
333	Blank	393	NeuroD1	453	Smad 5	513	Positive 2c
334	Blank	394	Neuropilin-2	454	Smad 7	514	Blank
335	LBP	395	Neurturin	455	Smad 8	515	TREM-1
336	LECT2	396	NGF R	456	SMDF / NRG1Isoform	516	TROY / TNFRSF19
337	Lefty - A	397	NOV / CCN3	457	Soggy-1	517	TSG-6
338	Leptin R	398	NRG1 Isoform GGF2	458	Sonic Hedgehog (Shh N-terminal)	518	TSLP
339	Leptin (OB)	399	NRG1-alpha / HRG1-alpha	459	SPARC	519	TWEAK / TNFSF12
340	LFA-1 alpha	400	NRG1-beta1 / HRG1-beta1	460	Spinesin	520	TWEAK R / TNFRSF12
341	LIF	401	NRG2	461	TACI / TNFRSF13B	521	Ubiquitin+1
342	LIF R alpha	402	NRG3	462	Tarc	522	uPA
343	LIGHT / TNFSF14	403	NT-3	463	TCCR / WSX-1	523	uPAR
344	Lipocalin-1	404	NT-4	464	TECK / CCL25	524	Vasorin
345	LRP-1	405	Orexin A	465	TFPI	525	VCAM-1 (CD106)
346	LRP-6	406	Orexin B	466	TGF-alpha	526	VE-Cadherin
347	L-Selectin (CD62L)	407	OSM	467	TGF-beta 1	527	VEGF
348	Luciferase	408	Osteoactivin / GPNMB	468	TGF-beta 2	528	VEGF R2 (KDR)
349	Lymphotactin / XCL1	409	Osteocrin	469	TGF-beta 3	529	VEGF R3
350	Lymphotoxin beta / TNFSF3	410	Osteoprotegerin / TNFRSF11B	470	TGF-beta 5	530	VEGF-B
351	Lymphotoxin beta R / TNFRSF3	411	OX40 Ligand / TNFSF4	471	TGF-beta RI / ALK-5	531	VEGF-C
352	MAC-1	412	PARC / CCL18	472	TGF-beta RII	532	VEGF-D
353	MCP-1	413	PD-ECGF	473	TGF-beta RIIb	533	VEGI / TNFSF15
354	MCP-2	414	PDGF R alpha	474	TGF-beta III	534	WIF-1
355	MCP-3	415	PDGF R beta	475	Thrombopoietin (TPO)	535	WISP-1 / CCN4
356	MCP-4 / CCL13	416	PDGF-AA	476	Thrombospondin (TSP)	536	XEDAR
357	M-CSF	417	PDGF-AB	477	Thrombospondin-1	537	Neg
358	M-CSF R	418	PDGF-BB	478	Thrombospondin-2	538	Internal Control 2a
359	MDC	419	PDGF-C	479	Thrombospondin-4	539	Internal Control 2b
360	MFG-E8	420	PDGF-D	480	Thymopoietin	540	Internal Control 2c

## VII. Troubleshooting Guide

<b>Problem</b>	<b>Cause</b>	<b>Recommendation</b>
Weak signal or no signal	1. Taking too much time for Detection. 2. Film developer does not work properly. 3. Did not mix HRP-streptavidin well before use. 4. Sample is too dilute. 5. Other.	1. The whole Detection process must be completed in 30 min. 2. Fix film developer. 3. Mix tube containing HRP-Conjugate Streptavidin well before use since precipitates may form during storage. 4. Increase sample concentration 1. Reduce blocking concentration by diluting in 1X Wash Buffer II. 2. Slightly increase HRP concentrations. 3. Slightly increase biotinylate-antibody concentrations. 4. Expose film for overnight to detect weak signal.
Uneven signal	1. Bubbles formed during incubation. 2. Membranes were not completely covered by solution.	1. Remove bubbles during incubation. 2. Completely cover membranes with solution.
High background	1. Exposure to x-ray file is too long. 2. Membranes were allowed to dry out during experiment. 3. Sample is too concentrated.	1. Decrease exposure time. 2. Completely cover membranes with solution during experiment. 3. Use more diluted sample.

## VIII. Reference List

1. Profiling of cytokine expression by biotin-labeled-based protein arrays. Ying Lin, Ruochun Huang, Lipai Chen *et al.* **Proteomics**. 2003, 3: 1750-1757.
2. Proteomic profiling of the cancer microenvironment by antibody arrays. Vladimir Knezevic, Chidchanok Leethanakul, Verena E. Bichsel *et al.* **Proteomics** 2001, 1, 1271–1278.
3. Antibody microarray profiling of human prostate cancer sera: Antibody screening and identification of potential biomarkers Jeremy C. Miller, Heping Zhou, Joshua Kwekel, Robert Cavallo *et al.* The, Brian B. Haab. **Proteomics**. 2003, 3: 56-63.
4. Smad4 signalling in T cells is required for suppression of gastrointestinal cancer. Byung-Gyu Kim, Cuiling Li, Wenhui Qiao, Mizuko Mamura, Barbara Kasperczak, Miriam Anver, Lawrence Wolfram, Suntaek Hong, Elizabeth Mushinski, Michael Potter, Seong-Jin Kim, Xin-Yuan Fu, Chuxia Deng and John J. Letterio. **Nature**. 2006; Vol 441: 1015.
5. Connexin suppresses human glioblastoma cell growth by down-regulation of monocyte chemotactic protein 1, as discovered using protein array technology. Ruochun Huang, Ying Lin, Cheng C. Wang, Jacob Gano, Biaoyang Lin, Qian Shi, Alton Boynton, Jocelyn Burke, and Ruo-Pan Huang. **Cancer Res**. 2002; 62:2806-2812.
6. LPS induces the interaction of a transcription factor, LPS-induced TNF-a factor, and STAT6(B) with effects on multiple cytokines.

Xiaoren Tang, Deborah Levy Marciano, Susan E. Leeman and Salomon Amar. *PNAS*. April 5, 2005 vol. 102 no. 14, 5132-5137.

7. HIV-1-mediated apoptosis of neuronal cells: Proximal molecular mechanisms of HIV-1-induced encephalopathy. Yan Xu, Joseph Kulkosky, Roger J. Pomerantz. *PNAS*. 2004 May 4, 2004 Vol. 101 No. 18.
8. A novel method for high-throughput protein profiling from conditioned media and patient's sera. Ruo-Pan Huang, Ruochun Huang, Yan Fan, and Ying Lin. *Ana. Biochem.* 2001;294(1):55-62.

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