

10X Tricine Running Buffer

- 1. Catalog No.** KTR030
- 2. Quantity** 500 ml
- 3. Storage & Stability** Store at R.T.
- 4. Description** 10X Tricine Running Buffer is used for running Protein samples for SDS PAGE analysis on Polyacrylamide gels.
- 5. Recommended Loading** Add 1/10 volume of 10X Tricine Running Buffer to 9/10 volume of D.W.
- 6. Protocol**
1. Tricine Sample Buffer does not contain reducing agent. For reducing conditions, add 0.2ml of 2-Mercaptoethanol or 0.08g of DTT to 4ml of Tricine Sample Buffer (5X) before use.
 2. Add four part of sample to one part of Tricine Sample Buffer (5X) and mix well. Heat the sample at 85°C for 2 minutes.
 3. Prepare 800ml 1X Tricine Running Buffer by adding 80ml of Tricine Running Buffer (10X) to 720ml of deionized water before use. Fill the upper and lower buffer chambers of EzCell with the appropriate amounts of running buffer. Make sure that the upper running buffer covers the sample wells completely.
 4. Load sample into the wells. Typically 0.1-0.5ug of protein per band with Coomassie Blue staining will give optical band intensity.
 5. Run the gel according to the following running conditions.

Voltage	125V constant	
Approx. Current	Start	80mA / 1.0mm gel
	End	40mA / 1.0mm gel
Approx. Run Time	65 minutes	

Turn off the power when the CBB dye is migrated to the end of the gel.

6. After the run, remove the gel from the cassette.
7. Fix, stain or transfer as desired.

* Buffer composition

Tricine Running Buffer (10x)	
Tris base	1 M
Tricine	1 M
SDS	1 %
pH	8.3

For research use only; not for use as a diagnostic



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